



Ottawa Hull K1A 0C9

(21) (A1) 2,108,113  
(22) 1993/10/08  
(43) 1995/04/09

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(51) INTL.CL. C12N-015/52; C12N-015/76; C12N-009/00; C12P-017/18;  
C12N-001/21

(19) (CA) **APPLICATION FOR CANADIAN PATENT** (12)

(54) DNA Sequence Encoding Enzymes of Clavulanic Acid  
Biosynthesis

(72) Jensen, Susan E. - Canada ;  
Aidoo, Kwamena A. - Canada ;  
Paradkar, Ashish S. - Canada ;

(71) Governors of the University of Alberta (The) - Canada ;

(57) 39 Claims

5,095,2/24

Notice: This application is as filed and may therefore contain an  
incomplete specification.



Industrie Canada Industry Canada

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Canada

DNA SEQUENCE ENCODING ENZYMES OF CLAVULANIC ACID  
BIOSYNTHESIS

This invention relates to methods for the production  
5 of the antibiotic, clavulanic acid.

Background of the Invention

Clavulanic acid is a broad spectrum beta-lactamase  
inhibitor and is an important antibiotic for the  
10 treatment of infectious diseases. It is produced  
commercially by the gram-positive mycelial prokaryote  
Streptomyces clavuligerus, which also produces the  $\beta$ -  
lactam antibiotics penicillin N, desacetoxo  
cephalosporin C and cephamycin C. Until recently,  
15 however, the pathway employed for clavulanic acid  
biosynthesis was much less well understood than the  
pathways leading to these other antibiotics.

Without knowledge of the pathway for clavulanic acid  
biosynthesis, it was not possible to isolate the genes  
20 coding for the key enzymes and to manipulate these genes  
to increase antibiotic yield or permit production of the  
antibiotic in heterologous systems.

One of the earliest enzymes of the pathway to be  
purified and characterised was clavaminic acid synthase.  
25 Two isozymes have now been identified and characterised  
(Marsh et al., (1992), Biochem., vol. 31, pp. 12648-657).

European Patent Application 0349121 describes a DNA  
restriction fragment encoding a portion of the genetic  
information involved in clavulanic acid synthesis but  
30 provides no sequence information.

Until the work of the present inventors, the  
complete complement of genes required for clavulanic acid  
synthesis had not been identified. The present inventors  
have now isolated, cloned and sequenced an 11.6 kb  
35 genomic DNA sequence from S. clavuligerus which codes for  
eight proteins and enables the production of clavulanic

Figure 7 shows an alignment of the amino acid sequence of CLA (S. clavuligerus CLA) with those of E. Coli agmatine ureohydrolase (E. Coli AUH), yeast arginase (yeast ARG), rat arginase (rat ARG) and human arginase (human ARG).

Figure 8 shows a Southern blot of NcoI digests of genomic DNA from five presumptive mutants (lanes 1-5) and from wild-type S. clavuligerus (lane 6). Panel A : membranes probed with cla-specific probe. Panel B : membranes probed with tsr-specific probe.

Figure 9 shows restriction enzyme maps of S. clavuligerus DNA inserts in cosmids. A. Restriction enzyme map of cosmid K6L2. B. Partial restriction enzyme map of cosmid K8L2. C. Restriction map of cosmids K6L2 and K8L2 indicating location of pcbC gene in relation to cla. D. The 2.0 kb NcoI fragment encompassing the cla gene used in generating nested deletions for sequencing. Abbreviations: Ba, BamHI; B, BglIII; E, EcoR1; K, KpnI; N, NcoI; S, SalI; and Sm, SmaI.

Figure 10 shows the deduced amino acid sequence (Sequence ID No.:3) of ORF1 of Figure 2.

Figure 11 shows the deduced amino acid sequence (Sequence ID No.:4) of ORF2 of Figure 2.

Figure 12 shows the deduced amino acid sequence (Sequence ID No.:5) of ORF3 of Figure 2.

Figure 13 shows the deduced amino acid sequence (Sequence ID No.:6) of ORF4 of Figure 2.

Figure 14 shows the deduced amino acid sequence (Sequence ID No.:7) of ORF5 of Figure 2.

Figure 15 shows the deduced amino acid sequence (Sequence ID No.:8) of ORF6 of Figure 2.

Figure 16 shows the deduced amino acid sequence (Sequence ID No.:9) of ORF7 of Figure 2.

Figure 17 shows the deduced amino acid sequence (Sequence ID No.:10) of ORF8 of Figure 2.

Figure 18 shows the deduced amino acid sequence (Sequence ID No.:11) of ORF9 of Figure 2.

when introduced into the non-clavulanate producer S. lividans as described in Example 4, enabled that species to produce clavulanic acid. This indicates that the 11.6 kb fragment contains all the genetic information required for clavulanate production.

As will be understood by those skilled in the art, the identification of the DNA sequence encoding the enzymes required for clavulanate synthesis will permit genetic manipulations to modify or enhance clavulanate production. For example, clavulanate production by S. clavuligerus may be modified by introduction of extra copies of the gene or genes for rate limiting enzymes or by alteration of the regulatory components controlling expression of the genes for the clavulanate pathway.

Heterologous organisms which do not normally produce clavulanate may also be enabled to produce clavulanate by introduction, for example, of the 11.6 kb DNA sequence of the invention by techniques which are well known in the art, as exemplified herein by the production of S. lividans strains capable of clavulanate synthesis. Such heterologous production of clavulanic acid provides a means of producing clavulanic acid free of other contaminating clavams which are produced by S. clavuligerus.

Suitable vectors and hosts will be known to those skilled in the art; suitable vectors include pIJ702, pJOE829 and pIJ922 and suitable hosts include S. lividans, S. parvulus, S. griseofulvus, S. antibioticus and S. lipmanii.

Additionally, the DNA sequences of the invention enable the production of one or more of the enzymes of the clavulanate pathway by expression of the relevant gene or genes in a heterologous expression system.

The DNA sequences coding for one or more of the pathway enzymes may be introduced into suitable vectors and hosts by conventional techniques known to those skilled in the art. Suitable vectors include pUC118/119

7  
shown in Figure 3. ORF 4 corresponds to cla. ORF 1, 7 & 8 are oriented in the opposite direction to pcbC. ORFs 2-6 and ORF 10 are all oriented in the same direction as pcbC. ORFs 2 and 3, and ORFs 4 and 5 are separated by very short intergenic regions suggesting the possibility of transcriptional and translational coupling. Table 1 summarises the nucleotide sequences and lengths of ORFs 1-10.

When the predicted amino acid sequences of proteins encoded by ORFs 1 - 10 were compared to protein sequence databases, some similarities were noted in addition to the already mentioned similarity between CLA and enzymes of arginine metabolism. ORF 1 showed a low level of similarity to penicillin binding proteins from several different microorganisms which are notable for their resistance to  $\beta$ -lactam compounds.

An EcoRI fragment of the 15 kb DNA sequence, containing 11.6 kb DNA, was cloned into a high copy number shuttle vector and introduced into S. lividans, as described in Example 4. Of seventeen transformants examined, two were able to produce clavulanic acid, indicating that the 11.6 kb fragment contains all the necessary genetic information for clavulanic acid production.

This 11.6 kb fragment encompasses ORF 2 to ORF 9 of the 15 kb DNA sequence.

ORF 2 shows a high degree of similarity to acetohydroxyacid synthase (AHAS) enzymes from various sources. AHAS catalyses an essential step in the biosynthesis of branched chain amino acids. Since valine is a precursor of penicillin and cephamycin antibiotics, and valine production is often subject to feedback regulation, it is possible that a deregulated form of AHAS is produced to provide valine during the antibiotic production phase. Alternatively, an AHAS-like activity may be involved in clavulanic acid production. While the presently recognized intermediates in the clavulanic acid

EXAMPLESExample 1Bacterial strains, vectors and growth conditions.

- 5        Streptomyces clavuligerus NRRL 3585, Streptomyces  
jumonjiniensis NRRL 5741, Streptomyces lipmanii  
NRRL 3584, Streptomyces griseus NRRL 3851, Nocardia  
lactamdurans NRRL 3802 and Streptomyces cattleya NRRL  
3841 were provided by the Northern Regional Research  
10   Laboratories, Peoria, Il. Streptomyces antibioticus ATCC  
8663 and Streptomyces fradiae ATCC 19609 were obtained  
from the American Type Culture Collection, Rockville, MD.  
Streptomyces lividans strains 1326 and TK24 were provided  
by D.A. Hopwood (John Innes Institute, Norwich, U.K.),  
15   Streptomyces venezuelae 13s and Streptomyces griseofuscus  
NRRL B-5429 were obtained from L.C. Vining (Department of  
Biology, Dalhousie University, Halifax, N.S.). Cultures  
were maintained on either MYM (Stuttard (1982) J. Gen.  
Microbiol., v. 128, pp. 115-121) or on a modified R5  
20   medium (Hopwood et al. (1985) in "Genetic Manipulation of  
Streptomyces : a laboratory manual", John Innes  
Foundation, U.K.) containing maltose instead of glucose  
and lacking sucrose (R5-S). Escherichia coli MV1193  
(Zoller and Smith (1987) Methods in Enzymology, v. 154,  
25   pp. 329-349), used as recipient for all of the cloning  
and subcloning experiments, was grown in Luria Broth (LB;  
Sambrook et al. (1989) in "Molecular Cloning : a  
laboratory manual", Cold Spring Harbour, N.Y.) or on LB  
30   agar (1.5%) plates containing ampicillin (50 µg/mL) or  
tetracycline (10 µg/mL). The cloning vectors pUC118 and  
pUC119 (Vieira and Messing (1987) Methods in Enzymology,  
v. 153, pp. 3-11) were provided by J. Vieira (Waksman  
Institute of Microbiology, Rutgers University,  
Piscataway, N.J.). The plasmid vector pJOE829 was  
35   generously provided by J. Altenbuchner (University of  
Stuttgart, Stuttgart, Germany). The plasmid pIJ702 was  
obtained from the American Type Culture Collection,

The probe was designed as an 8-fold degenerate mixture of oligonucleotides to take into consideration the biased codon usage of Streptomyces (Bibb et al., 1984, Wright and Bibb (1992), Gene, v. 113, pp. 55-65).).

- 5 End-labelled probe was then used to screen a cosmid library of S. clavuligerus genomic DNA fragments as described in Materials and Methods.

A library of S. clavuligerus genomic DNA fragments (15-22 kb size fractionated fragments) was constructed as  
10 previously described (Doran et al. (1990), J. Bacteriol., v. 172, pp. 4909-4918). using the cosmid vector pLAFR3. A collection of 1084 isolated E. coli colonies containing recombinant cosmids was screened for the presence of cla using the 24-mer mixed oligonucleotide probe (Fig. 1)  
15 which had been end-labelled with [ $\gamma$ -<sup>32</sup>P]dATP and polynucleotide kinase (Boehringer Mannheim). Colony hybridization and subsequent washing was performed as described by Sambrook et al., (1989), at 55°C with a final wash in 0.2X SSC (1X SSC, 0.15M NaCl and 0.015M  
20 sodium citrate) and 0.1% SDS.

Five colonies which gave strong hybridization signals were isolated from the panel of 1084 clones, and restriction analysis showed that the positive clones contained overlapping fragments of DNA. Two clones, K6L2  
25 and K8L2, with sequences that spanned about 40 kb of the S. clavuligerus genome, were chosen for further analysis. Clone K8L2 contained about 22 kb of S. clavuligerus genomic DNA and included a portion of cla and all of the pcbC gene which encodes IPNS in the penicillin/cephamycin biosynthetic pathway. A restriction map of K6L2 is shown  
30 in Fig. 9. Within the approximately 27 kb of DNA contained in K6L2, the oligonucleotide probe hybridized to a 2.0 kb NcoI fragment which was subsequently found to contain the entire cla gene. Hybridization studies,  
35 restriction mapping and DNA sequence analysis revealed that cla was situated 5.67 kb downstream of the pcbC gene of S. clavuligerus (Fig. 9).

program described above. The AUH sequence had previously been aligned with the three ARG sequences (Szumanski & Boyle (1990), J. Bacteriol., v. 172, pp. 538-547). Identical matches in two or more sequences are indicated with upper case letters.

### Example 2

#### DNA hybridization

Genomic DNA preparations from various Streptomyces species were isolated as described by Hopwood et al. (1985). For interspecies DNA hybridization analysis, 2.0 µg amounts of genomic DNA preparations were digested with NcoI for 16h, and electrophoresed in 1.0% agarose gels. The separated DNA fragments were then transferred onto nylon membranes (Hybond-N, Amersham) and hybridized with a cla specific probe prepared by labeling an internal 459 bp SalI fragment (Fig. 1) with [ $\alpha$ -<sup>32</sup>P]dATP by nick translation. Hybridization was done as described by Sambrook et al., (1989). Hybridization membranes were washed twice for 30 min in 2X SSC; 0.1% SDS and once for 30 min in 0.1X SSC; 0.1% SDS at 65°C.

#### Sequences homologous to cla in other Streptomycetes

Three of six producers of  $\beta$ -lactam antibiotics, S. clavuligerus, S. lipmanii and S. jumonjinensis showed positive hybridization signals whereas S. cattleya, S. griseus, and N. lactamdurans did not (data not shown). None of the nonproducing strains examined, S. venezuelae, S. lividans, S. fradiae, S. antibioticus and S. griseofuscus gave any signal. All of the streptomycetes that gave positive signals were producers of clavam-type metabolites (Elson et al., 1987)

### Example 3

#### Disruption of the genomic cla gene

A 2.0 kb NcoI fragment that contained the entire cla gene was digested at its unique KpnI site and the ends



bioassay procedures described previously (Jensen et al. (1982), supra).

5 All of the resulting colonies with disrupted cla genes grew equally well on minimal medium and complex media and produced as much penicillin and cephamycin as did the wild-type, but produced no clavulanic acid (data not shown). HPLC analysis of cell supernatants confirmed the inability of the disrupted cla mutants to synthesize any clavulanic acid (data not shown).

10

#### Example 4

#### Protoplast formation and transformation

E. coli competent cell preparation and transformation were as described by Sambrook et al., (1989). Protoplasts of S. clavuligerus were, prepared, transformed and regenerated as described by Bailey et al. (1984), Bio/Technology, v. 2, pp. 808-811, with the following modifications. Dextrin and arginine in the regeneration medium were replaced by starch and sodium glutamate respectively. Protoplasts were heat shocked at 43°C for 5 min prior to the addition of DNA. Standard procedures were used for protoplasting and transformation of S. lividans (Hopwood et al. (1985)).

25 The 11.6 kb EcoRI fragment from K6L2 (Fig. 9) was cloned into the EcoRI site of pCAT-119. pCAT-119 is derivative of pUC119 which was prepared by insertionally inactivating the ampicillin resistance gene of pUC119 by the insertion of a chloramphenicol acetyltransferase gene (Jensen et al. (1989), Genetics & Molec. Biol. of Ind. Microorg., pp. 239-245 Ed. Hersherberger, Amer. Soc. Microbiol). The PCAT-119 plasmid carrying the 11.6 kb fragment was then digested with PstI and ligated to the Streptomyces plasmid pIJ702, which had also been digested with PstI. The resulting bifunctional plasmid carrying 35 the 11.6kb insert was capable of replicating in either E. coli (with selection for chloramphenicol resistance) or in S. lividans (with selection for thiostrepton

Example 5Sequencing of 15 kb DNA fragment

Ordered sets of deletions were generated as described in Example 1 using fragments of the DNA insert from the cosmid clone K6L2 (Figure 9) and subcloned into the E. coli plasmids pUC118 and pUC119. Overlapping fragments were chosen which extended from the end of the pcbC gene downstream for a distance of about 15 kb ending at the BglIII site. The deletion generated fragments were sequenced in both orientations as described in Example 1. The sequence is shown in Figure 2.

The present invention is not limited to the features of the embodiments described herein, but includes all variations and modifications within the scope of the claims.

The embodiments of the invention in which an exclusive property or privilege is claimed are defined as follows:

- 5 1. An isolated genomic DNA molecule comprising the nucleotide sequence of Figure 2 (Sequence ID No.:1).
2. An isolated DNA molecule having the nucleotide sequence of nucleotides 2033 to 13636 of Figure 2  
10 (Sequence ID No.:20).
3. An isolated DNA molecule having the nucleotide sequence of nucleotides 109 to 1764 of Figure 2 (Sequence ID No.:21).
- 15 4. An isolated DNA molecule having the nucleotide sequence of nucleotides 2216 to 3937 of Figure 2 (Sequence ID No.:22).
- 20 5. An isolated DNA molecule having the nucleotide sequence of nucleotides 3940 to 5481 of Figure 2 (Sequence ID No.:23).
- 25 6. An isolated DNA molecule having the nucleotide sequence of nucleotides 5654 to 6595 of Figure 2 (Sequence ID No.:24).
7. An isolated DNA molecule having the nucleotide sequence of nucleotides 6611 to 7588 of Figure 2  
30 (Sequence ID No.:25).
8. An isolated DNA molecule having the nucleotide sequence of nucleotides 7895 to 9076 of Figure 2 (Sequence ID No.:26).

35

18. An isolated DNA molecule comprising a nucleotide sequence encoding the amino acid sequence of Figure 15.
- 5 19. An isolated DNA molecule comprising a nucleotide sequence encoding the amino acid sequence of Figure 16.
- 10 20. An isolated DNA molecule comprising a nucleotide sequence encoding the amino acid sequence of Figure 17.
- 15 21. An isolated DNA molecule comprising a nucleotide sequence encoding the amino acid sequence of Figure 18.
- 20 22. An isolated DNA molecule comprising a nucleotide sequence encoding the amino acid sequence of Figure 19.
23. An isolated protein having the amino acid sequence of Figure 10.
24. An isolated protein having the amino acid sequence of Figure 11.
- 25 25. An isolated protein having the amino acid sequence of Figure 12.
- 30 26. An isolated protein having the amino acid sequence of Figure 13.
27. An isolated protein having the amino acid sequence of Figure 14.
- 35 28. An isolated protein having the amino acid sequence of Figure 15.

transforming the host with a DNA molecule comprising a nucleotide sequence encoding one or more of the enzymes of the clavulanate synthetic pathway.

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FIGURE 2 - 1

	1	10	20	30	40	50	60	
1	gcggaaccgg	ccgcccctga	gcggggcggc	cggggaaggaa	acgggcccgt	cgccccctcg	60	
61	ggagggggcg	gccggcccgt	ccggtgcgcg	cggtgggtgc	ggcgcgggTC	AGCCGGCCGC	120	End of ORF 1
121	GAGGTTGCTG	AGGAACTTCG	CGCGGACGGG	GCCCGCGTCG	GCGCCGCCCG	ACCCGCCGTC	180	
181	CTCCAGCAGG	ACCGACCAGG	CGATGTTCCG	GTGCCCCTGG	TAGCCGATCA	TCCAGGCGTG	240	
241	CGTCTTCGGC	GGCTTCTCGG	TGCCGAAGTC	GGCGGTACCG	GTCTTGCGGT	GCGGCTGTCC	300	
301	GCCGAGGCC	CGCAGGGCGT	CGCCGGCGCC	GTGCGTGACG	GTGGAACGCA	TCATGGAACG	360	
361	CAGCGAGTCG	ACGATGCCCC	GGGCCATCCG	GGGGGCTGG	TGCGGCTTCT	TGACCGCGTC	420	
421	GGGCACCAGC	ACGGGCTGCT	TGAAGTCGCC	CTGCTTGACG	GTGGCGGCGA	TGGAGGCCAT	480	
481	CACCAGGGGC	GACGCCTCGA	CCCTGGCCTG	TCCGATGGTG	GACGCGGCCT	TGTCGTTCTC	540	
541	GCTGTTGGAG	ACGGGGACGC	TGCCGTCGAA	GGTGGAGGCG	CCGACGTCCC	AGGTGCCGCC	600	
601	GATGCCGAAG	GCTTCGGCGG	CCTGCTTCAG	GCTGGACTCG	GAGAGCTTGC	TGCGGGAGTT	660	
661	GACGAAGAAC	GTGTTGCAGG	AGTGGGCGAA	GCTGTCCCGG	AAGGTCGAGC	CCGCGGGCAG	720	
721	CGTGAAGTGG	TCCTGGTTCT	CGAAGCTCTG	GCCGTTGACA	TGGGCGAACT	TCGGGCAGTC	780	
781	GGCCCGCTCC	TCCGGGTTCA	TCCCCTGCTG	GAGCAGGGCC	GCGGTGGTGA	CCACCTTGAA	840	
841	GGTGGAGCCG	GGCGGGTAGC	GGCCCTCCAG	CGCGCGGTTT	ATGCCGGAGG	GCACGTTTCG	900	
901	GGCGGCCAGG	ATGTTGCCGG	TGGCGGGGTC	GACGGCGACG	ATCGCCGCGT	TCTTCTTCGA	960	
961	GCCCTCCAGG	GCCGCCGCGG	CGGCGGACTG	GACCCGCGGG	TCGATGGTGG	TCTTCACCGG	1020	
1021	CTTGCCCTCG	GTGTCTTGA	GGCCGGTGAG	CTTCTTGACC	ACCTGGCCGG	ACTCACGGTC	1080	
1081	CAGGATCACG	ACCGAGCGCG	CCGCGCCGGA	GCCGCCGGTG	AGCTGCTTGT	CGTAGCGGGA	1140	
1141	CTGGAGGCC	GCCGAGCCCT	TGCCGGTCCT	GGGGTCGACC	GCGCCGATGA	TGGAGGCGGC	1200	
1201	CTGGAGGACA	TTGCCGTTGG	CGTCGAGGAT	GTCCGCGCGC	TCCGCGACT	TGAGGGCGAG	1260	
1261	GGTCTGCCCC	GGAACCATCT	GCGGATGGAT	CATCTCGGTG	TTGAACGCGA	CCTTCCAATC	1320	
1321	CTTGCCGCCG	CCGACGACCT	TCGCGGTGGA	GTCCAGGCG	TACTCCCCGG	CCCCGGGGAG	1380	
1381	GGTCATTCTG	ACGGTGAACG	GTATCTCCAC	CTCGCCCTCG	GGGTTCTTCT	CCCCGGTCTT	1440	
1441	GGCGGTGATC	TCCGTCTTCG	TCGGCTTGAG	GTTGGTCATG	ACGGATTTGA	TCAGCGACTC	1500	
1501	GGCGTTGTCC	GGGGTGTCG	TCAGCCCGGC	GGCCGTCGGG	GCGTCGCCCT	TCTCCAGGC	1560	

*Sim, M. Baum*

FIGURE 2 - 3

3241 CGTGGAGCAC TTCGAGACCG CGACCGCCTC CTTGCGGGCC AAGCAGCGCC ACGACATCGA 3300  
 3301 GCCGCTGCGC GCCCGGATCG CGGAGTTCTT GGCCGACCCG GAGACCTACG AGGACGGCAT 3360  
 3361 GC GCGTCCAC CAGGTCATCG ACTCCATGAA CACCGTCATG GAGGAGGCCG CCGAGCCCGG 3420  
 3421 CGAGGGCACG ATCGTCTCCG ACATCGGCTT CTTCCGTAC TACGGTGTGC TCTTCGCCCC 3480  
 3481 CGCCGACCAG CCCTTCGGCT TCCTCACCTC GGCGGGCTGC TCCAGCTTCG GCTACGGCAT 3540  
 3541 CCCCCCGCC ATCGGCGCCC AGATGGCCCG CCCGGACCAG CCGACCTTCC TCATCGCGGG 3600  
 3601 TGACGGCGGC TTCCACTCCA ACAGCTCCGA CCTGGAGACC ATCGCCCGGC TCAACCTGCC 3660  
 3661 GATCGTGACC GTCGTCGTC ACAACGACAC CAACGGCCTG ATCGAGCTGT ACCAGAACAT 3720  
 3721 CGGTCACCAC CGCAGCCACG ACCCGGGGGT CAAGTTCGGC GCGCTCGACT TCGTCGCGCT 3780  
 3781 CGCCGAGGCC AACGGTGTG ACGCCACCCG CGCCACCAAC CGCGAGGAGC TGCTCGCGGC 3840  
 3841 CCGTGC AAG GGTGCCGAGC TGGGTCTGTC GTTCTCATC GAGGTCCCG TCAACTACGA 3900  
 3901 CTTCCAGCCG GCGGGCTTCG GCGCCCTGAG CATCTGATCA TGGGGGCACC GGTCTTCCG 3960  
 3961 GCTGCCITCG GGTTCCTGGC CTCCGCCCGA ACGGGCGGGG GCCGGGCCCC CGGCCCGGTC 4020  
 4021 TTCGCGACCC GGGGCAGCCA CACCGACATC GACACGCCCC AGGGGGAGCG CTCGCTCGCG 4080  
 4081 GCGACCCTGG TGCACGCCCC CTCGGTCGCG CCCGACCGCG CGGTGGCGCG CTCCTCACC 4140  
 4141 GGCGCGCCCA CCACCGCGGT GCTCGCCGGT GAGATCTACA ACCGGGACGA ACTCCTCTCC 4200  
 4201 GTGCTGCCCC CCGGACCCGC GCCGAGGGG GACGCGGAGC TGGTCTGCG GCTGCTGGAA 4260  
 4261 CGCTATGACC TGCATGCCTT CCGGTGGTG AACGGGCGCT TCGCGACCGT GGTGCGGACC 4320  
 4321 GGGGACCGGG TCCTGCTCGC CACCGACCAC GCCGGTTCGG TGCCGCTGTA CACCTGTGTG 4380  
 4381 GCGCCGGGCG AGGTCCGGGC GTCCACCGAG GCCAAGGCGC TCGCCGCGCA CCGCGACCCG 4440  
 4441 AAGGGCTTCC CGCTCGCGGA CGCCCGCCGG GTCGCCGGTC TGACCGGTGT CTACCAGGTG 4500  
 4501 CCCGCGGGCG CCGTGATGGA CATCGACCTC GGCTCGGGCA CCGCGTCAC CCACCGCACC 4560  
 4561 TGGACCCCGG GCCTCTCCCG CCGCATCCTG CCGGAGGGCG AGGCCGTGCG GGCCGTGCGG 4620  
 4621 GCCGCGCTGG AGAAGGCCGT CGCCAGCGG GTCACCCCG GCGACACCCG GTTGGTGGTG 4680  
 4681 CTCTCCGGCG GAATCGACTC CTCCGGGGTC GCGGCCTGTG CGCACCGGGC GGCCGGGGAA 4740  
 4741 CTGGACACGG TGTCCATGGG CACCGACACG TCCAACGAGT TCCGCGAGGC CCGGGCGGTC 4800  
 4801 GTCGACCATC TGCGACCCG GCACCGGGAG ATCACCATCC CGACCACCGA GCTGCTGGCG 4860

*Sim, M. Bunn*

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FIGURE 2 - 5

6541 GATCGGTGCG GAACTGCTCT ACCAGTACGC CCGAGCCCCAC <sup>End of ORF 4--></sup> AGAACCCAGT TGTGAoggg 6600  
 6601 acatcgtgtc <sup>Beginning of ORF 5--></sup> ATGGCCTCTC CGATAGTTGA CTGCACCCCG TACCGCGACG AGCTGCTCGC 6660  
 6661 GCTCGCCTCC GAGCTTCCCG AGGTGCCGCG CGCGGACCTC CATGGCTTCC TCGACGAGGC 6720  
 6721 GAAGACGCTG GCCGCCCGTC TCCCGGAGGG GCTGGCCGCC GCTCTCGACA CCTTCAACGC 6780  
 6781 CGTGGGCAGC GAGGACGGTT ATCTGCTGCT GCGCGGGCTG CCCGTCGACG ACAGCGAGCT 6840  
 6841 GCCCAGAGACG CCGACCTCCA CCCC GGCCCC GCTGGACCGC AAGCGGCTGG TGATGGAGGC 6900  
 6901 CATGCTCGCG CTGGCCGGCC GCCGGCTCGG TCTGCACACG GGGTACCAGG AGCTGCGCTC 6960  
 6961 GGGCACGGTC TACCACGACG TGTACCCGTC GCCCGGCGCG CACTACCTGT CCTCGGAGAC 7020  
 7021 CTCCGAGACG CTGCTGGAGT TCCACACGGA GATGGCGTAC CACATCCTCC AGCCGAACCTA 7080  
 7081 CGTCATGCTG GCCTGCTCCC GCGCGGACCA CGAGAACCGG GCGGAGACGC TGGTCGGCTC 7140  
 7141 GGTCCGCAAG GCGCTGCCCC TGCTGGACGA GAAGACCCGG GCCCGTCTCT TCGACCGCAA 7200  
 7201 GGTGCCCTGC TGCCTGGACG TGGCCTTCCG CGGCGGGGTC GACGACCCGG GCGCGATCGC 7260  
 7261 CAACGTCAAG CCGCTCTACG GGGACGCGAA CGACCCGTTT CTCGGGTACG ACCGCGAGCT 7320  
 7321 GCTGGCGCCG GAGGACCCCG CGGACAAGGA GGCCGTCGCC CATCTGTCCC AGGCGCTCGA 7380  
 7381 CGATGTGACC GTCGGGGTGA AGCTCGTCCC CGGTGACGTC CTCATCATCG ACAACTTCCG 7440  
 7441 CACCACGCAC GCGCGGACGC CGTTCTCGCC CCGCTGGGAC GGGAAAGGACC GCTGGCTGCA 7500  
 7501 CCGCGTCTAC ATCCGCACCG ACCGCAATGG ACAGCTCTCC GGCGGCGAGC GCGCGGGCGA 7560  
 7561 CACCATCTCG <sup>End of ORF 5--></sup> TTCTGCGCCG GCCGCTGAGc ccggctcccc gaggccctgg gccccggcgc 7620  
 7621 cgggaaccggc tcccggctct gccccctcac ccgcccgcgc ggtgaggggg caggccccctt 7680  
 7681 tgtgccgggt gccgtgcgtc ctgcgaggggt gccggggcgc gggggacggc ggaggtgccc 7740  
 7741 ggcggccggg tgccgtgcgc cgcctgtggg tgctgtacag cactccgtgt gccgtgcgc 7800  
 7801 accccgtgca taaatttgcc actctatggg aaataatgca gagtgcgacg ggtgagggcg 7860  
 7861 tcgcccgtgcc ctttcgtga caggagacgc <sup>Beginning of ORF 6--></sup> tgacATGTCC GACAGCACAC CGAAGACGCC 7920  
 7921 CCGGGGATTC GTGGTGCACA CGGCGCCGGT GGGCCTGGCC GACGACGGCC GCCACGACTT 7980  
 7981 CACCGTCTCT GCCTCCACCG CCCC GGCCAC CGTGAGCGCC GTCTTCACCC GCTCCCGCTT 8040  
 8041 CGCCGGGCGG AGCGTCGTGC TGTGCCGGGA GGCGGTGGCC GACGGGCAGG CGCGCGGTGT 8100  
 8101 GGTGGTGCTG GCCCGCAACG CGAATGTCGC GACCGGCCTG GAGGGCGAGG AGAACGCGCG 8160

Sim; M. Bunnif



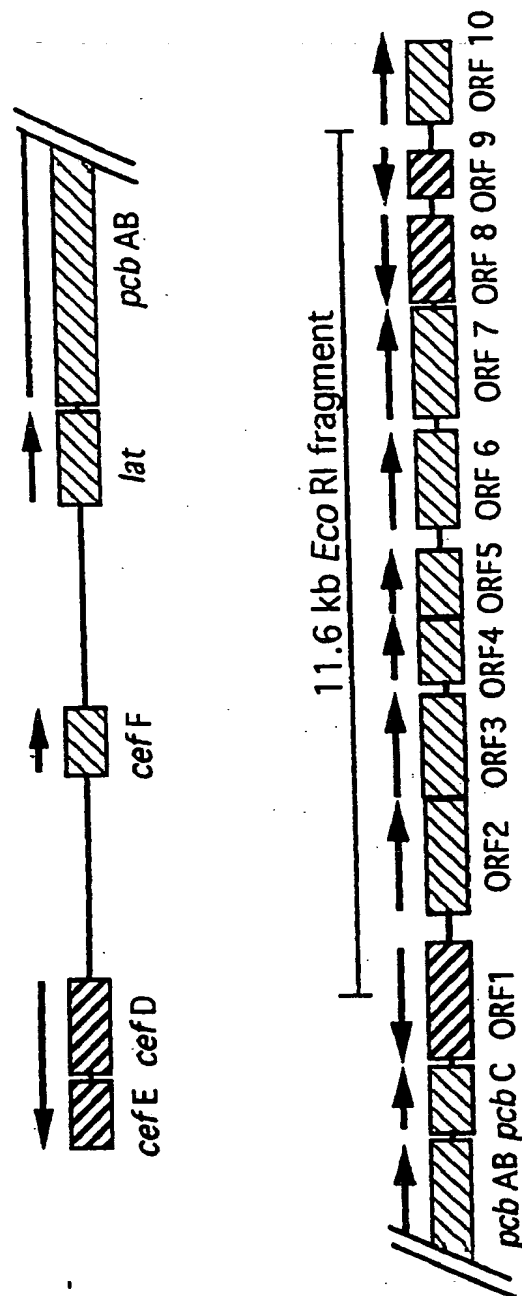
## FIGURE 2 - 7

9841 TACCGGCTGC GGCCCGTGGC GACCGGCCCG TACCGGATCG TCTCGTACAC CCGGGGCGAG 9900  
 9901 CTGGCCGTCC TGGAGCCCAA TCCGCACTGG GACCCCGAGA CCGACCCGGT GCGCGTCCAG 9960  
 9961 CGCGCCTCCC GGATCGAGGT GCACCTCGGC AAGGACCCGC ACGAGGTGGA CCGCATGCTG 10020  
 10021 CTGGCGGGCG AGGCCCATGT GGACCTCGCG GGCTTCGGTG TGCAGCCCGC GGCCAGGAG 10080  
 10081 CGCATCCTCG CCGAGCCGGA GCTGCGCGCG CACGCGGACA ACCCGCTGAC CGGCTTCACC 10140  
 10141 TGGATCTACT GCCTGTCGAG CCGGATCGCC CCGTTCGACA ATGTGCACTG CCGGCGGGCC 10200  
 10201 GTGCAGTTCG CCACCGACAA AGCGGCCATG CAGGAGGCGT ACGGCGGCGC GGTGGGCGGC 10260  
 10261 GACATCGCGA CCACCCTGCT GCGCCCGACC CTCGACGGCT ACAAGCACTT CGACCGCTAC 10320  
 10321 CCGGTCGGCC CCGAGGGCAC CGGCGACCTG GAGGCCGCC GCGCCGAGCT GAAGCTGGCC 10380  
 10381 GGGATGCCCG ACGGCTTCCG CACCAGGATC GCCGCCGCA AGGACCGGCT CAAGGAGTAC 10440  
 10441 CGGGCCGCCG AGGCGCTGGC CGCCGGGCTC GCCCGGGTCG GCATCGAGGC GGAGGTGCTG 10500  
 10501 GACTTCCCGT CGGGCGACTA CTTGACCGC TACGGCGGCT GCCCGGAGTA TCTGCGCGAG 10560  
 10561 CACGGGATCG GGATCATCAT GTTCGGCTGG GCGCGCGACT TCCCCGACGG ATACGGCTTC 10620  
 10621 CTCCAGCAGA TCACCGACGG GCGCGCGATC AAGGAGCGCG GCAACCAGAA CATGGGCGAG 10680  
 10681 CTGGACGACC CGGAGATCAA CGCGCTGCTG GACGAGGGGG CGCAGTGCGC CGACCCGGCG 10740  
 10741 CGGCGCGCGG AGATCTGGCA CCGCATCGAC CAGCTCACGA TGGACCACGC GGTATCGTT 10800  
 10801 CCGTATCTGT ACCCGCGGTC CCTGCTCTAC CGGCACCCGG ACACCCGCAA CGCCTTCGTC 10860  
 10861 ACCGGCTCCT TCGGGATGTA CGACTACGTG GCGCTCGGCG CGAAGTGAgc acgggggtccg 10920  
 10921 gccccgggac cgtatgtccc ggggcccggac cccgcccgtt ccccgcccgg tccgggtccgg 10980  
 10981 acccggtcgc ggcccgcTCA GCCGGACATC CCGGCCCCGG CCGCGACCCC GCGCCGGATC 11040  
 11041 GGCCAGTGGC CCTGCGCCAG GGGCCGTTCC ACGCTGCGGC AGGCGAGAGC GGCTTCGCGG 11100  
 11101 AACTCCGCTT CGTACAGCGC GAGCTGGCGC AGGAACTGCC GGGTCGGGCC GGTGAGGCTG 11160  
 11161 GTCCCCCGCG GGCTGCGCAG CAGCAGCCGG GCGCCGAGGG ACTGCTCCAG CCGGTGAATC 11220  
 11221 CGGCGGGTGA GCGCCGACTG GCTGATCGAC AGCACCGCCG CGGCCCGGTT GATGCTGCCG 11280  
 11281 TGCCGGGCCA CGGCCTGGAG CAGATGGAGA TCGTCCACAT CCAGTTTGC GGCCTCGGCC 11340  
 11341 TGGCCGGGCA CGGAGCCCTG GTCGGGTCCC GCCCCGAAGC GCGGGGCGTC CGCGCCGGTG 11400  
 11401 CGCTCCGCGT ACCACTGCGC CCACCAGGGC TCGTCCAGCA GGTGCGGGTG GTGTTGGCG 11460  
 11461 AAGCGCCGGA GCTGGACCTC GCGATCAGC GCGGCCAGCC GTCCCGCCAG CGCCCGGGC 11520

*Sim, M. Bunn*

13201 CCCGGCGGCG GTCAGCTCGT CACCCAGGGC GCGCAGCTTC TCGACCCGGC GCGCGGCGAT 13260  
13261 GGCCACGGCG GCGCCCTCGG CGGCCAGGGC GCGGGCCGTG GCCTCGCCGA TGCCCCAGCT 13320  
13321 CGCGCCCGTG ATGAGCGCGA CTTTCCCTG <sup>beginning of ORF 9</sup> GAGTGGGAT GGCATcattt cctccacatg 13380  
13381 gtgctgcat cgtggtgagc gtatgaagaa ggggtgagac ctgccgtgcc ggggcgggtt 13440  
13441 ccgtacgccg gaccgttgcg gtgggcacgg ccgaccgggt acggatggcc gcagttcccc 13500  
13501 ggggagttcc cggggaatgg tgaataccgc ggcctctcc gatggtcttc ggaggacacc 13560  
13561 cggggattca ccgggaatca gcggccggag ttctccccgt ccacggcaga cgctatcagc 13620  
13621 gtcgattcc ccggtgaatt cccttcgggtg gaccgggtta tgactgttcc gcgccgggtta 13680  
13681 tgcgcgcgc cccggcgagc cggccaccgc cccgggggct gcggcagatt gggcgccacg 13740  
13741 acatggcgcg agcagcgatc ggcgggtgAT <sup>Beginning of ORF 10---</sup> GATGAACGAG GCAGCGCCTC AGTCCGACCA 13800  
13801 GGTGGCACCG GCGTATCCGA TGCACCGGGT CTGCCCCGGT GACCCGCCGC CGCAACTGGC 13860  
13861 CGGGCTGCGG TCCCAGAAGG CCGCGAGCCG GGTGACGCTG TGGGACGGCA GCCAGGTGTG 13920  
13921 GCTGGTGACC TCGCACGCCG GGGCCCCGGC CGTCTGGGC GACCGCCGCT TCACCGCGGT 13980  
13981 GACGAGCGCG CCCGGCTTCC CGATGCTGAC CCGCACCTCC CAACTGGTGC GCGCCAACCC 14040  
14041 GGAGTCGGCG TCGTTCATCC GCATGGACGA CCCGCAGCAC TCCGGGCTGC GCTCGATGCT 14100  
14101 CACCCGGGAC TTCCTGGCCC GCCGCGCCGA GGCCTGCGC CCCGCGGTGC GGGAGCTGCT 14160  
14161 GGACGAGATC CTGGGCGGGC TGGTGAAGGG GGAGCGGCCG GTCGACCTGG TCGCCGGACT 14220  
14221 GACGATCCCG GTGCCCTCGC GGGTCATCAC CCTGCTCTTC GGCGCCGGTG ACGACCGCCG 14280  
14281 GGAGTTCATC GAGGACCGCA GCGCGGTCTT CATCGACCGC GGCTACACCC CGGAGCAGGT 14340  
14341 CGCCAAGGCC CGGGACGAAC TCGACGGCTA TCTGCGGGAG CTGGTCGAGG AGCGGATCGA 14400  
14401 GAACCCGGGC ACCGACCTGA TCAGCCGGCT CGTCATCGAC CAGGTGCGGC CGGGGCATCT 14460  
14461 GCGGGTCGAG GAGATGGTCC CGATGTGCCG GCTGCTGCTG GTGGCCGGTC ACGGCACCAC 14520  
14521 CACCAGCCAG GCGAGCCTGA GCCTGCTCAG CCTGCTCACC GACCCGGAGC TGGCCGGGCG 14580  
14581 CCTCACCAG GACCCGGCCC TGCTGCCCAA GGCGGTCGAG GAGCTGCTGC GCTTCCACTC 14640  
14641 CATCGTGCAG AACGGGCTGG CCCGTGCCGC GGTGGAGGAC GTCCAGCTCG ACGATGTGCT 14700  
14701 CATCCGGGCG GCGGAGGGCG TGGTGCTGTC GCTGTGCGCG GGCAACCGGG ACGAGACGGT 14760  
14761 CTCCCCGAC CCGGACCGGG TGGACGTGGA CCGCGACGCC CGCCGCCATC TCGCCTTCGG 14820  
14821 CCACGGCATG CACCAGTGCC TGGGCCAGTG GCTGGCCCCG GTGGAGCTGG AGGAGATCCT 14880

Sam, M. B.



ORF 4 = *cla*

FIGURE 3

*Simon, M. Barry*

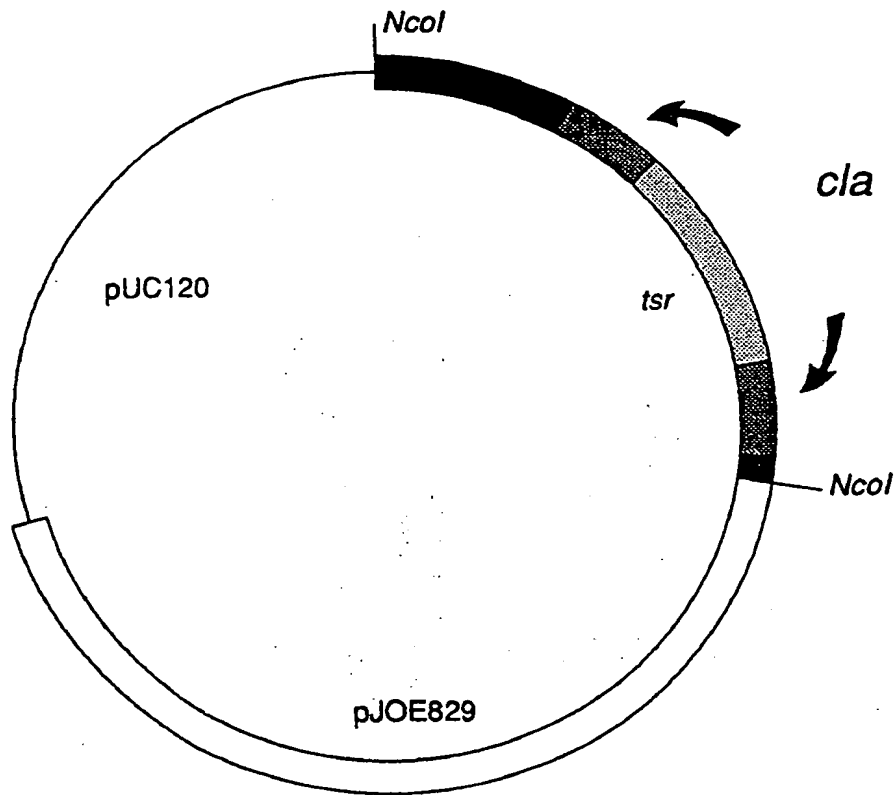


FIGURE 5

Sim; M. Barry

60  
 S. Cl. CLA 1 veridshvspryaqiptFmRLPhdpQPrgyDV--VvIGaPyDggTSyRpGARfGPqAIR  
 E. co. AUH MSTIGHqYdNslvSnafGFIRLPmnfQPydsDadwVitGvPfDmaTSgRaGGRhGPqAIR  
 yeast ARG MeT-GphY-NyyKnRelslvIAPFSgGQgkIGVEKGPKymIKhGL-qtsiedlgwsteLE  
 rat ARG MS-----sKpkpleIIGAPFSKGQPRGGVEKGPaaLRKAGL-----VE  
 human ARG MS-----aKSRTIGIIGAPFSKGQPRGGVEeGPTvLRKAGL-----LE

120  
 S. Cl. CLA 61 seSglihgvgidRgPgtFDI---INcYDaGDINItpfDmniaidtaQsHISgLLKANaaf  
 E. co. AUH qvStnl-awehnRfPwnFDmrrerINVVDcGDIVyafgDarEmSEKLQAhaeKLLaAGkrm  
 yeast ARG psmdeo-qfVgKIkmeKdsttggssVmidGVKakRadIVGEAtkIvynsYSKVvqANRfp  
 rat ARG KLKEtE-ynV-rDhGDLafvDvPNDSPFQIVKNPRS--VGKAnEQLAAvVAetaKNGtIS  
 human ARG KLKEqE-cdV-KDyGDLpFaDIPNDSPFQIVKNPRS--VGKASEQLagkVaqVKKNGRIS

180  
 S. Cl. CLA 121 LmIGGDHSLTvaalRAVAeqhGpLAVVHIDAHsDTNpafyGgryhHGTpFrhgideKLID  
 E. co. AUH LsfGGDHfvTIpILRAhAkHfGkmALVHfDAHTDTyan--GcefdHGTMFytapKEgLIID  
 yeast ARG LtLGGDHSIAIGtvSAVIDkyPDaGLIWIDAHaDINTi--esTpsGNLHGcPVsFLmgln  
 rat ARG vVLGGDHSmaIGSISsHARVHPDLcVIWVDAHTDINTP--LTTsSGNLHGQPVaFLLKEL  
 human ARG LVLGGDHSIAIGSISgHARVHPDLGVIWVDAHTDINTP--LTTsSGNLHGQPVsFLLKEL

240  
 S. Cl. CLA 181 PaamVQIGIRGHNPKPDSLdyarghGvrVvtAdefgelgVggtadLirekV-----  
 E. co. AUH PnhsvQIGIRt-----efdkdnGftVIdAcqvnDrsVddvIaqykqiy-----  
 yeast ARG KdvphcpesIk-----WVpgnISpKkIaYIGLRDvDaGEkkILKdLGLaaFSMyhVD  
 rat ARG KGKfPDVPGFS-----WVTPCISAKDIVYIGLRDvDPGEHYIITLGIKYFSMTEVD  
 human ARG KGKIPDVPGFS-----WVTPCISAKDIVYIGLRDvDPGEHYIITLGIKYFSMTEVD

300  
 S. Cl. CLA 241 -----GqRPVYvsVDIDvVDPAPAGTGTpPapGGLISREvLaLIR  
 E. co. AUH -----GdmPVYLtFDIDcLDPAFAPGTGTpVIGGLISdraikLVR  
 yeast ARG KyGlnaVIEmamkavhpoteGegPImcSyDVGVDPIyIPATGTpVRGGLTIREGLfLVE  
 rat ARG KLGIKVM-----ETfSYLLGRKKRPiHLSFDVDGLDPvFTPATGTpVVGGLsYREGLYITE  
 human ARG rLGIGKVM-----ETfSYLLGRKKRPiHLSFDVDGLDPsFTPATGTpVVGGLTYREGLYITE

360  
 S. Cl. CLA 301 cv-gDLkpVGfDVMEVsPIYDhggITSi-----IATeIgaELLYqyArahrTqIz  
 E. co. AUH gL-KDLNIVGmDVVEVaPaYDaseITaI-----AAAtIAEmLYIaAaKkge  
 yeast ARG rLaesGNLlaLDVVEcNPdLaihdIhYsnTisagcAIArcALGetII  
 rat ARG EIYKTGLLSGLDIMEVNPTLGKTPEEVTRTVNTAVpITLSCFGtkREGNHKPoDYlKPPK  
 human ARG EIYKTGLLSGLDIMEVNPTLGKTPEEVTRTVNTAVAITLACFGLaREGNHKP-IDYLnPPK

FIGURE 7

*Sim; M. Baum*

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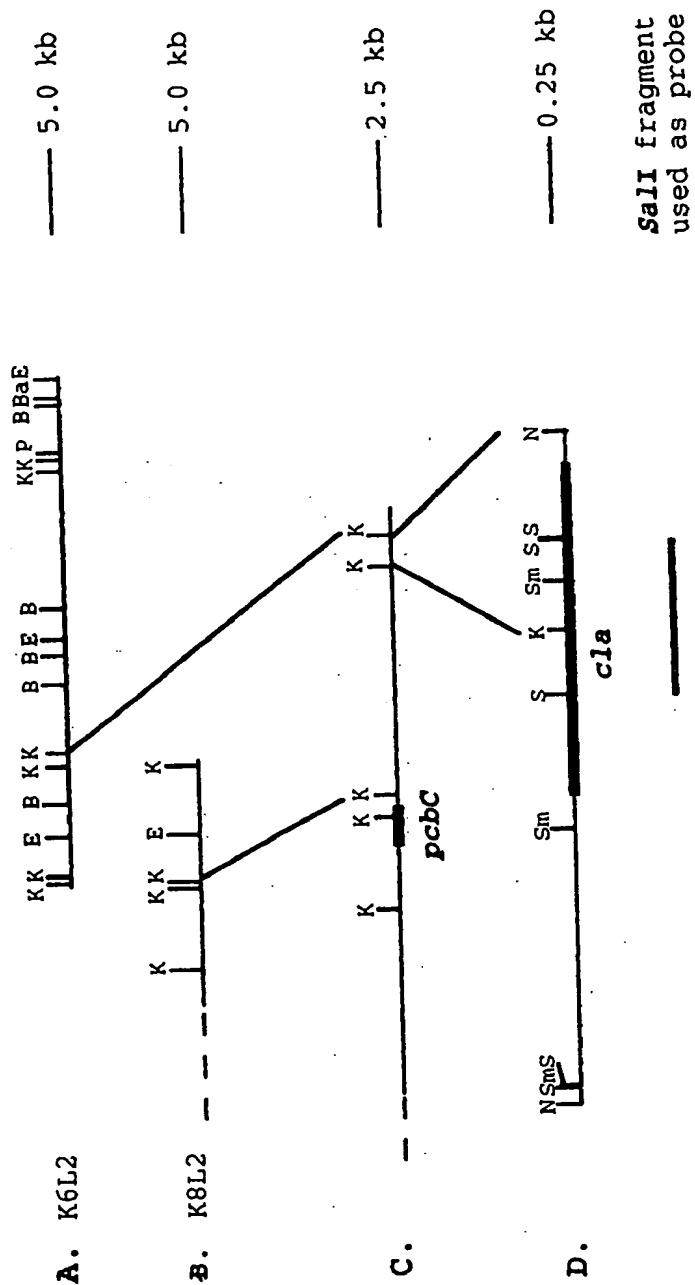


FIGURE 9

Sim; M. Baum

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	10	20	30	40	50	60	
1	MSRVSTAPSG	KPTAAHALLS	RLRDHGVGKV	FGVVGREAAS	ILFDEVPID	FVLTRHEFTA	60
61	GVAADVLARI	TGRPQACWAT	LGPGMTNLST	GIATSVLDRS	PVIALAAQSE	SHDIFPNDTH	120
121	QCLDSVAIVA	PMSLYAVELQ	RPHEITDLVD	SAVNAAMTEP	VGPSFISLPV	DLGSGSEGID	180
181	TTVPNPPANT	PAKPVGVVAD	GWQKAADQAA	ALLAEAKHPV	LVVGAAAIRS	GAVPAIRALA	240
241	ERLNIPVITT	YIAKGVLPVG	HELVYGAVTG	YMDGILNFPA	LQTMFAPVDL	VLTVGYDYAE	300
301	DLRPSMWQKG	IEKKTVRISP	TVNPIPRVYR	PDVDVVTDL	AFVEHFETAT	ASFGAKQRHD	360
361	IEPLRARIAE	FLADPETYED	GMRVHQVIDS	MNTVMEEAAE	PGEPTIVSDI	GFFRHYGVLF	420
421	ARADQPFGL	TSAGCSSFGY	GIPAAIGAQM	ARPDQPTFLI	AGDGGFHSNS	SDLETIARLN	480
481	LPIVTVVWNN	DTNGLIELYQ	NIGHHRSHDP	AVKFGGVDFV	ALAEANGVDA	TRATNREELL	540
541	AALRKGAELG	RPFLIEVPVN	YDFQPGGFGA	LSIZ			574
	10	20	30	40	50	60	

FIGURE 11

*Sim; of. Lunny*

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	10	20	30	40	50	60	
1	VERIDSHVSP	RYAQIPTFMR	LPHDPQPRGY	DVVVIGAPYD	GGTSYRPGAR	FGPQAIRSES	60
61	GLIHGVGIDR	GPGTFDLINC	VDAGDINLTP	FDMNIAIDTA	QSHLSGLLKA	NAAFLMIGGD	120
121	HSLTVAALRA	VAEQHGPLAV	VHLDHSDTN	PAFYGGRYHH	GTPFRHGIDE	KLIDPAAMVQ	180
181	IGIRGHNPKE	DSL DYARGHG	VRVVTADDFG	ELGVGGTADL	IREKVGQRPV	YVSVDIDVVD	240
241	PAFAPGTGTP	APGGLLSREV	LALLRCVGDL	KPVGFDVMEV	SPLYDHGGIT	SILATEIGAE	300
301	LLYQYARahr	TQLZ					314
	10	20	30	40	50	60	

FIGURE 13

*Sim; M. Barry*



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	10	20	30	40	50	60	
1	MSDSTPKTPR	GFVVHTAPVG	LADDGRHDFT	VLASTAPATV	SAVFTRSRFA	GPSVVLCREA	60
61	VADGQARGVV	VLARNANVAT	GLEGEENARE	VREAVARALG	LPEGEMLIAS	TGVIGRQYPM	120
121	ESIREHLKTL	EWPAGEGGFD	RAARAIMTTD	TRPKEVRVSV	GGATLVGIAK	GVGMLEPDMA	180
181	TLLTFFATDA	RLDPAEQDRL	FRRVMDRTFN	AVSIDTDTST	SDTAVLFANG	LAGEVDAGEF	240
241	EEALHTAALA	LVKDIASDGE	GAAKLIEVQV	TGARDDAQAK	RVGKTVVNSP	LVKTAVHGCD	300
301	PNWGRVAMAI	GKCSDDTDID	QERVITIRFGE	VEVYPPKARG	DQADDALRAA	VAEHLRGDEV	360
361	VIGIDLAIAD	GAFTVYGCDL	TEGYVRLNSE	YTTZ			394
	10	20	30	40	50	60	

FIGURE 15

*Simon, M. Barry*

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	10	20	30	40	50	60	
1	MEVARRTGVR	HGTVERRLDR	LDRIVGLPLT	LSRHTARLT	TAGSRILVAG	RRFFHQVDLA	60
61	ARTHIFGHGS	EAVDAPEVLS	LVSTEPLLDE	VVEDAAASLD	LLLSVRHEAP	HQVAAQLAGY	120
121	QVDAAYTWSL	QSPRHSLEERS	VRTCEVLDDP	LWVILPRDHP	LAARREVSLA	DLRDETWSSE	180
181	TGPGSEILVT	RVFQLAGLTA	PTRLHITGAS	VARGILRRGD	AIGLGSPTHP	AVQDPSLVRR	240
241	SLAERPRRTT	SLLVDPTIVP	RALAGRLAAL	IAEVQLRRFA	EHHRDLLDEP	WWAQWYAERT	300
301	GADARRFGAG	PDQGSVPGQA	EGRKLDVDDL	HLLQAVARHG	SINRAAAVLS	ISQSALTRRI	360
361	HRLEQSLGAR	LLLRSPRGTS	LTGPTROFLR	QLALYEAEFR	EAALACRSVE	RPLAQGHWPI	420
421	RRGVAAGARM	SGZ					433
	10	20	30	40	50	60	

FIGURE 17

*Sim; M. Bunn*

	10	20	30	40	50	60	
1	MMNEAAPQSD	QVAPAYPMHR	VCPVDPPPQ	AGLRSQKAAS	RVTLWDGSQV	WLVTSAGAR	60
61	AVLGDRRFTA	VTSAPGFPM	TRTSQVLVRAN	PESASFIRMD	DPQHSRLRSM	LTRDFLARRA	120
121	EALRPVAVREL	LDEILGGLVK	GERPVDLVAG	LTIPVPSRVI	TLLFGAGDDR	REFIEDRSV	180
181	LIDRGYTPEQ	VAKARDEL	YLRELVEERI	ENPGTDLISR	LVIDQVRPGH	LRVEEMVPMC	240
241	RLLLVAGHGT	TTSQASLSLL	SLLTDPELAG	RLTEDPALLP	KAVEELLRFH	SIVQNGLARA	300
301	AVEDVQLDDV	LIRAGEGVVL	SLSAGNRDET	VFPDPDRVDV	DRDARRHLAF	GHGMHQCLGQ	360
361	WLARVELEEI	LA AVL RWMPG	ARLAVPFEEL	DFRHEVSSYG	LGALPVTWZ		409
	10	20	30	40	50	60	

FIGURE 19

*Simon; M. Baum*

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